Stockroom Recipes

Back to; the Stockroom, the Biology Department, or the College

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These recipes were collected over the course of many years by various stockroom technicians (therefore I can take no credit or responsibility for their accuracy or lack thereof). For each recipe there are surely other ways of making them and/or other formulations. Some are actually formulas concocted by our faculty for their own research needs.

There are many duplications in this list (ie. rat ringers is also listed as ringers, rat etc.) as I have intended it to be easily downloaded and printed to a card type file with cross references. This is most easily done by "searching" for the ****** lines and "replacing" with a page break. The resulting pages should fit easily on a 4x6 cards.

Should you wish to add or correct errors in this list please e-mail your recipe or correction to me, Bob McNulty and I will incorporate it/them into our list with the appropriate acknowledgements (if desired).

BEFORE MIXING ANY REAGENT YOU SHOULD UNDERSTAND THE PROPERTIES OF ALL OF THE CHEMICALS INVOLVED.

Consult your MSDS, Merk Index, CRC etc. before mixing any reagent.

The Biology Department, California State University Chico, and most important, Myself.... assume NO responsibility for the outcome of mixing any of these reagents !!!!!

Now on with the list

To avoid laborious scrolling click on the desired location in the alphabet below.

<u>A B C D E F G H I J K L M N O P Q R S T U V W X Y Z</u>

A's

2-4 - Dichlorophenoxyacetic acid (2-4-D)

Mix dry reagent in a small amount of 95% ethanol. Place this solution in a warming plate 32 deg C and add prewarmed water slowly with swirling until desired concentration is reached.

5% Ammoniacal Silver Nitrate

Add concentrated ammonium hydroxide to 5% silver nitrate until resulting precipitate again dissolves.

A.F.A. , Botanical

115 ml Formalin 115 ml Glacial acetic acid 805 ml 95% Ethanol 805 ml Distilled water

ACD used for blood

Na Citrate 1.32 gm 0.48 gmCitric Acid 1.47 gm Dextrose 100 mls Distilled water

Use 0.25 ml/ml of blood

ACETIC ACID - DILUTIONS

172 MLS 57.3 mls 1N 1.046 mls 0.02N Bring to a final volume of 1 Liter.

Aceto Carmine Stain (Stern)

45 ml Glacial acetic acid

55 ml water 0.5 gm Carmine

Heat the 45% acetic acid solution to boiling. Add the 0.5gm

Carmine and continue heating for several minutes with

stirring. Filter and cool.

ACETO-CARMINE (BELLING'S)

STOCK 1

55 ml Distilled water Acetic acid 45 ml 5.0 gms Carmine

STOCK 2 55 ml

Distilled water 45 ml Acetic Acid 5 gms Ferric oxide (iron acetate)

Boil stock solution 1 for 15 minutes, cool and filter To 50 mls of stock 1 add stock 2 until pcpt just appears

then quickly add 50 ml more of stock 1

***for most purposes omit stock solution 2 and just use stock solution 1.

ACETO-CARMINE (chromosome stain for drosophila)

- Weigh sufficient solid Carmin or Orcein, or Lacmoid to make a 2% solution.
- Place the stain with the appropriate volume of 45% acetic acid in a flask with a vertical condensor.
- 3. Reflux all day, glass beads or pumice will help to prevent bumping.
- 4. Allow to cool and filter.

ACETOCARMINE

Stock sol'n - Boil and excess (aprox 0.5g/100 ml) of carmine in 45% acetic acid (aq) 2-4 minutes. Cool and filter. Working sol'n. Dilute stock sol'n with 45% acetic acid 1:2. An iron mordanted stain often is favored because of darker bluish tinged red. Belling added a few drops of 50% ferric hydrate in 50% acetic acid, but only a few drops! Too much iron will produce a pcpt in a short time. Moree(1944) determined quantatively the amount of ferric chloride to add and includes volumes tables Moree, r. 1944 Stain tech 9:103-108

ACETIC ALCOHOL

100 mls Ethanol anhydrous 33 ml Glacial Acetic Acid

ACID ALCOHOL

50 ml 35% Ethanol 3 drops HCl (con)

ACID ALCOHOL FOR GLASSWARE CLEANING

100ml 70% Ethanol

5 ml HCl

ACID CLEANING SOLUTION

120 gms Na Dichromate 1 Liter Tap Water 1.6 Liters Con Sulfuric

Dissolve chromate in water. Add acid with swirling. Be

sure to surround mixing vessel with ice bath.

Acid cleaning solution 1

120 gm Na dichromate 1000 ml tap water 1600 ml sulfuric acid

Dissolve chromate in water. Add acid with swirling. Be

sure to surround mixing vessel with ice bath.

ACID CLEANING SOLUTION 2

120 gm Potassium Dichromate

600 mls water

920 mls Con Sulfuric Acid

Dissolve chromate in water. Add acid with swirling. Be

sure to surround mixing vessel with ice bath.

Acid cleaning solution 2

120 gm Potassium dichromate

600 ml water

920 ml sulfuric acid

Dissolve chromate in water. Add acid with swirling. Be

sure to surround mixing vessel with ice bath.

AFA (ALEXANDER'S)

460 ML FORMALIN 1152 ML 95% ETHANOL

76 ML GLACIAL ACETIC ACID

2308 ML TAP WATER

ALCOHOLIC BORAX CARMINE (GRENACHER'S)

3 gm Carmine 4 gm Borax

100 ml distilled water

Boil until the carmine is dissolved $(1/2\ hr)$ or better yet allow the mixture to stand at room temp

until dissolved.

Then add 70% ethanol (100 ml) and allow to stand

for a day or two then filter.

ALEXANDER'S AFA

460 ML FORMALIN 1152 ML 95% ETHANOL

76 ML GLACIAL ACETIC ACID

2308 ML TAP WATER

ALGAL PRESERVATIVE

8 parts 50% Ethanol 1 part 5% Formalin 1 part Glycerine

Alizerine Red S for whole mount staining of bone

1 part alizerin red in 10,000 parts 1% - 2% KOH Material is cleared in 2% KOH first.

Alkaline - Iodine

NaOH 500 gm

кон 700 gm

*******and********
NaI 135 gm
or

KI 150 gm

dissolved in distilled water to make 1 Liter (used for determining dissolved oxygen)

Alkaline Alcohol

Add a few drops of 1.0% sodium bicarbonate to 70% Ethanol

Alkaline Na I

Dissolve 40 gm of NaOH and 90 gm of NaI to make 100 ml of solution using distilled water.

Alum Carmine (Grenacher's)

100 ml Distilled water 3 gm Aluminum ammonium

Combine and boil for 15 mon or until carmine is

dissolved. Cool and filter.

Alum Cochineal

6 gm Potassium alum 6 gm powdered cochineal 90 ml distilled water

Boil for 1/2 hour. After the fluid has settled, decant the supernatant liquid, add more water and boil it down until only 90 mls remain. Filter when cool and add a bit of thymol or a little salicylic acid to prevent

the growth of mold

Aman's Lactophenol

20 ml melted phenol
20 ml Lactic acid
40 ml glycerine
21 ml distilled water

To 100 ml of the above add 5 ml of 1% aqueous Cotton Blue

or Acid Fuchsin.

Aminonaphthal Sulfonic Acid

Grind 0.5 gm of 1-amino-2-naphthal-4-sulfonic acid with 5 ml of sodium sulfite solution containing 1 gm anhydrous sodium sulfite per 5 ml and disolve in 200 ml sodium bisulfite solution containting 30 gm Na2S2O5 per 200 ml.

Keep in brown tightly stoppered bottle.

Ammonium Hydroxide Concentrations

Reagent grade (58%) = 29.7 N U.S.P. grade (28%) = 15 N

Ammonium Potassium Oxalate

1.2 gm Ammonium oxalate
0.8 gm Potassium oxalate
100 ml distilled water

Aqua Regia

1 part Nitric Acid

3 parts HCl

1 part distilled water

WARNING!!! IF YOU DON'S ADD THE WATER POISONOUS

CHLORINE GASS WILL FORM!!!!!!!!!

B's

<u>A B C D E F G H I J K L M N O P Q R S T U V W X Y Z</u>

Balsam in 100% Ethanol

1 part Balsam (solid)
1 part absolute ethanol

This takes at least 3 days to disolve so prepare in

advance.

Barfoed's Solution

Place 4.5 gm crystalized neutral cupric acetate in 100 ml of water, add 0.12 ml of 50% acetic acid.

Baritt's Reagent test for acetylmethylcarbinol

5% solution of alpha naphthol in absolute ethyl alcohol. 40% solution of potassium hydroxide.
0.3% creatin in 40% potassium hydroxide but ADD JUST BEFORE USE. Stable for 2 weeks.

Baryta Water

Add Barium Hydroxide plus a pinch of Barium Chloride to a bottle fo distilled water until no more will go into solution.

Filter and keep tightly stoppered.

BELLING'S ACETO - CARMINE

STOCK 1

55 ml Distilled water 45 ml Acetic acid 5.0 gms Carmine

STOCK 2

55 ml Distilled water
45 ml Acetic Acid
5 gms Ferric oxide
(iron acetate)

Boil stock solution 1 for 15 minutes, cool and filter To 50 mls of stock 1 add stock 2 until pcpt just appears

then quickly add 50 ml more of stock 1

***for most purposes omit stock solution 2 and just use stock solution 1.

Benedict's Reagent

173 gm Sodium Citrate
100 gm Sodium Carbonate
(anhydrous)
17.3 gm Copper Sulfate
to make

to make

1 liter Distilled water

Dissolve sodium citrate and sodium carbonate in 800 ml of distilled water. Filter and add copper sulfate

dissolved in 100 ml of distilled water. Add distilled water to make 1 Liter.

Benzidine Reagent

Make a saturated solution of benzidine in glacial acetic acid.

Benzidine Test

Place a drop of blood in 3 ml of water. Add 2 ml of saturated solution of benzidine in glacial acetic acid. To this add 1 ml of 3% hydrogen peroxide.

Berlese's Medium

10 ml water

3 ml glacial acetic acid 5 ml dextrose syrup 8 gm gum acacia 75 gm Chloral Hydrate

Mix water, acid, and syrup. Dissolve gum acacia in this mixture (may take a week). Stir at intervals. Be careful not to include any air bubbles. Afeter gum acacia is

dissolved, a;dd chloral hydrate.

Biuret Reagent tried and true!!!!

3 gm CuSO4*5H2O 12 gm Potassium sodium tartrate 600 ml 10% NaOH (aq)

Dissolve copper sulfate in 500 ml of distilled water, then dissolve potassium sodium tartrate in this solution. Boil 800 mls of distilled water, cool covered. Add the 600 mls of 10% NaOH to the cooled water.

Mix the two solutions and bring to 2 liters. Bottle in

PLASTIC BOTTLES.

Biuret Reagent and Paper

Biuret Reagent is prepared by adding 25 ml of 3% Copper Sulfate in 10% Potassium hydroxide. Paper (Walkers modification)

Add 1% copper sulfate solution a drop at a time, with stirring to a 40% solution of NaOH unitl the mixture becomes a deep blue color. Then filter paper may be immersed in the reagent, dried, and cut into strips for use in tests for proteins.

Blood Diluting Fluid (Gower's)

12.5 gm Sodium Sulfate 33.3 ml Glacial acetic acid 200 ml distilled water

Borax Carmine, Alchohlic, Grenacher's

3 gm Carmine Borax

distilled water

Boil until the carmine is dissolved (1/2 hr) or better yet allow the mixture to stand at room temp until dissolved. Then add 70% ethanol (100 ml) and allow to stand for a day or two then filter.

Botanical F.A.A.

115 ml Formalin

115 ml Glacial acetic acid 805 ml 95% Ethanol 805 ml Distilled water

Bouin's Fixative (Alcoholic) Kistner

150 ml 90% Ethanol 60 ml Formalin

15 ml Glacial acetic acid

1 gm Picric Acid *****************

Bouin's Picro-Formol Fixative

75 parts saturated aq Picric

Acid

25 parts Formalin

5 parts Glacial acetic Acid

1 gm of picric acid will saturate about 75 mls of water.

Bradford Protein Assay Dye Reagent

50 mg Coomassie Blue G250

100 ml 85% H3PO4 850 ml H2O 50 ml methyl alcohol

Mix commassie blue in methyl alcohol first. Then add this to the phosphoric acid. Now add 500 mls of water, mix and filter. Add and mix the remaining 350 mls of water

bottle and store in refrigerator.

Brazil's Fixative

150 ml 80% Ethanol 1 gm Picric Acid 60 ml Formalin

15 ml Acetic acid ADD JUST BEFORE USE

Brodie's Manometer Fluid

23 gm NaCl

5 gm Sodium Choleate 500 ml distilled water

density = 1.033, Po = 10000

Evans blue $(200\,\mathrm{mg/L})$ or acid fuchsin are excellent dyes for the fluid; other dyes may be used, but some of these tend to decompose in the manometer. Determine the density of the solution with a pycnometer.

Brom Cresol Purple - indicator for fermentation

1.6 gm $$\operatorname{Brom}$ Cresol Purple Mix BCP in dry morter with 18.5 ml of).1N NaOH. Dillute to 100 ml (Enough additional to completely disolve

the reagent)

Brom Thymol Blue - indicator

0.1 gm Brom Thymol Blue
16.0 ml 0.01N NaOH
Mix in a mortar and then dillute to 250 ml.
Use 5 drops / 10 ml of solution being checked.

Brom Thymol Blue - indicator

0.4 gm Brom Thymol Blue 500 ml 95% Ethanol 500 ml Distilled water

Dissolve the indicator in the alcohol. Add the distilled

water, stir and then filter through fast paper.

Budding Yeast

Place a small portion of a yeast cake in malt extract broth Incubate while shaking.

Buffer Solutions of Clark and Lubs - Group 1

To 50 ml of a 0.2M KCl add the indicated number of mls of a 0.2N HCl solution and then diltue to $200\,\mathrm{mls}$ with distilled water.

INDICATOR: Thymol Blue

Buffer Solutions of Clark and Lubs - Group 2

To 50 ml of a 0.2M Potassium acid bi phthalate add the indicated number of mls of 0.2N HCl and dilute to 200 ml with distilled water $\,$

INDICATOR: Thymol blue and Brom Phenol blue

Buffer Solutions of Clark and Lubs - Group 3

To 50 ml of a 0.2M Potassium acid bi phthalate add the add the indicated number of mls of 0.2N NaOH, and dilute with distilled water to 200 ml.

INDICATORS: Brom cresol green, Brom phenol Blue, Brom
 cresol purple

Buffer Solutions of Clark and Lubs - Group 4

To 50 ml of a 0.2M Acid Potassium phosphate add the indicated number of ml of 0.2M NaOH and dillute to 200 ml. INDICATORS: Brom thymom blue, Brom cresol blue, and Phenol red

Buffer Solutions of Clark and Lubs - Group 5

To 50 ml of 0.2N Boric acid in 0.2M KCl add the indicated number of ml of 0.2N NaOH and diltue to 200 ml with distilled water.

INDICATORS: Cresol red and thymol blue

Buffer, Phosphate, Sorensen's

Stock solution a) = 9.08gm / 1000ml water of KH2-PO4 b) = 9.47gm / 1000ml water of Na2-HPO4 Combine the two stock solutions as indicated

Buffer, Tris-Maleate

Dissolve 29 gm of maleic acid and 30.3 gm of tris-hydroxymethyl aminomethane in 500 ml of water. Add 2 g of Charcoal, shake and let stand for 10 min. Filter. Mix 40 ml of this stock solution with the amount of 1N NaOH indicated below and dillute to a final volume of 100 ml using water.

C's

 $\underline{A} \quad \underline{B} \quad \underline{C} \quad \underline{D} \quad \underline{E} \quad \underline{F} \quad \underline{G} \quad \underline{H} \quad \underline{I} \quad J \quad \underline{K} \quad \underline{L} \quad \underline{M} \quad \underline{N} \quad O \quad \underline{P} \quad Q \quad \underline{R} \quad \underline{S} \quad \underline{T} \quad U \quad V \quad \underline{W} \quad X \quad \underline{Y} \quad \underline{Z}$

Carbol Methyl Green

1 gm Methyl green

5 gm Phenol or Carbolic

10 ml 95% ethanol 90 ml water

Put phenol in mixture o 10 ml alcohol and 60 ml of water. Heat without boiling. Dissolve phenol with constant stirring. Put methyl green in 30 ml of water and heat. Pour dye solution into phenol solution (DO NOT REVERSE THIS! Let stand overnight. Decant and filter.

Carboxylene

Melt some Carbolic acid crystals (Phenol) over a low flame (avoid breathing fumes). Mix one part acid to three parts Xylene.

Carl's Solution (FAA)

170 ml 95% ethanol 60 ml Formalin

20 ml Glacial acetic acid

280 ml water

Add the glacial acetic acid to the solution just

before using.

Carmine, Alum, Grenacher's

100 ml distilled water 3 gm aluminum ammonium

sulfate Lgm Carmine

Combine and boil for 15 min or until carmine is dissolved.

Cool and filter.

Carmine, Semicon's

1 part glacial acetic acid

l part water

Add carmine powder in excess of that which dissolves immediately. Heat to 95-100 deg C for 15 min. Cool and filter. Dilute filtrate eith equal partsw of 70% ethanol

Carnoy's Fixative

6 parts 100 % ethanol 1 part glacial acetic acid

3 parts Chloroform

Carnoy-LeBrun Fluid

30 ml 100% ethanol

30 ml glacial acetic acid

30 ml Chloroform

aprox 10gm Mercuric Chloride Saturat mercuric chloride in the solution.

Celestine Blue (Gray's)

Dissolve 2.5 gm iron alum in 100 ml of water with 14 ml glycerine. Place 1 gm Celestine Blue B in a beaker. Tilt the beaker and tap to accumulate the dye in one place. Pour on 0.5 ml con sulfuric acid and mix with glass rod. When the effervescence has ceased the dye will be in the form of a friable mass. Break up this mass coursly and pour on, with constant stirring, the soltuion of inom alum heated to 50 deg C. Cool to room temp and adjust to pH 0.8 with sulfuric acid. Allow to settle for several days and do not stir when removing some stain for specimens.

Cerophyl Infusion for Stock Culture

4.7 ml .1M Na2-HPO4 0.3 ml .1M NaH2-PO4 945 ml distilled water

50 ml ballanced salt sol'n

0.5 gm Cerophyl

Add cerophyl to about 200 ml of boiling water and boil for

3 min. Add remaining water, buffer and pour into

appropriate vessels. BALANCED SALT SOLUTION =

distilled water 100 ml 2.08 gm NaCl MgS04 0.08 gm 0.17 gm MgCl2 CaCl2 0.02 gm KCl 0.46 gm

Chromate Cleaning Solution 1

120 gm Na dichromate 1000 ml tap water 1600 ml sulfuric acid

Dissolve chromate in water. Add acid with swirling. Be

sure to surround mixing vessel with ice bath.

Chromate Cleaning Solution 2

120 gm Potassium dichromate

600 ml water

920 ml sulfuric acid

Dissolve chromate in water. Add acid with swirling. Be

sure to surround mixing vessel with ice bath.

Chromate Starch

5 gm Potassium dichromate 1 gm soluable starch

100 ml water

Suspend the starch and the potassium dichromate in cold water with stirring. Bring the mixture to a gentle boil while stirring, cool.

Chromatographic solvent, alkaline butanol sat HOH

80 ml n- butanol

20 ml amonium hydroxide

sp gr 0.90 100 ml water

Shake together vigorously and allow to stand.

Chromosome stain for fruit fly salivary glands

Weigh sufficient solid Carmin or Orcein, or Lacmoid to make a 2% solution.

- Place the stain with the appropriate volume of 45% acetic acid in a flask with a vertical condensor.
- Reflux all day, glass beads or pumice will help to prevent bumping.
- Allow to cool and filter.

Cleaning solution for blood diluting pipets

Pull the following solutions through the pipet in order.

Never let blood dry in the pipet

- 1. Dilute acetic acid (4-5 %)
- 2. tap water
- 3. Distilled water

- 4. Acetone
- 5. Air until enclosure bounces free

Clearing Solution for leaves pg 1 of 2

- A. Pre treatment
- Dried soak in water till material sinks.
 Fresh extract chlorophill rinse in water.
- B. Clearing
- 1. transfer tislsue to a vial of 5% aq solution of NaOH, hydrolize and remove cytoplasimic material.
- 2. if delicate, clear at room temp, otherwise heat may be used - 55 deg C. Replace with fresh NaOh when discolored
- 3. if not cleared in several days wash thoroughly in water and soak in 10% clorox for 10 - 20 minutes.

Clearing Solution for leaves pg 2 of 2

- 4. when tissue is milky white, wash thouroughly in HOH
- C. Stainging
- 1. 1% analine blue in 95% ethanol
- 2. 1% safranin in 50% ethanol
- 3. 0.5% fast green in 100 % ethanol
- 4. 0.5% crystal violet in 100 % ethanol

Cobalt Chloride weather paper

A piece of white blotting paper is immersed in a solution containing two parts cobalt chloride to one part comon salt. While wet the pater will remain pink, but when dried in the sun or near a bunsen burner, it turns blue.

This is the basis of the weather pictures sold in the shops. A homemade one works just as well. A picture containing sky or water dcan be cut from a book and an inset of this prepared blotting paper made to replace the sky. Mount the picture on a card and hang near a window wher it will quickly respond to changes in the hygrometric state of the atmosphere.

Colchicine Solution

0.0125 gm Colchicine

Coliform test broth

10 gm Protease Peptone #3

1 gm Beef extract 5 gm Lactose

Sodium Chloride 5 gm 18 mg Phenol red

1 liter water

Cotton blue

20 ml carbolic acid 20 ml Lactic acid 40 ml glycerine 20 ml water 0.05 gm Cotton blue

Crystral violet 1:1000

0.9 gm Crystal Violet (85% dye content)

100 mls Water ******************

Cyanophycean Medium

KNO3 0.1 gm K2-HPO4 0.05 gmMgSO4*7HOH

10 drops 1% Ferric ammonium

citrate

The above should be added to 1000ml of distilled deionized

This can be solidified by adding agar to reach a 1.5%

concentration and heating to dissolve it.

D's

 $\underline{C} \quad \underline{D} \quad \underline{E} \quad \underline{F} \quad \underline{G} \quad \underline{H} \quad \underline{I} \quad \underline{J} \quad \underline{K} \quad \underline{L} \quad \underline{M} \quad \underline{N} \quad \underline{O} \quad \underline{P} \quad Q \quad \underline{R} \quad \underline{S} \quad \underline{T} \quad U \quad V \quad \underline{W} \quad X \quad \underline{Y} \quad \underline{Z}$

Dahl's Pollen Mounting Medium

16 parts 85% Lactic acid 2 parts N-Butyl alcohol 95% Ethanol 1 part 1 part Glacial Acetic Acid

1 part Glycerine

Desicote

5 ml Beckman Desicote 95 ml Benzene (thiophene

free)

Dichlorophenoxyacetic acid (2-4-D)

Mix dry reagent in a small amount of 95% ethanol. Place this solution in a warming plate 32 deg C and add prewarmed water slowly with swirling until desired concentration is reached.

Dichromate Cleaning Solution

120 gm Na Dichromate

1000 ml Water

1600 ml Con Sulfuric Acid (technical grade)

Digestive juice (synthetic)

7-10 ml con HCl

to 1 Liter Distilled water

6 gm Pepsin

Dilution Fluid, Turk's

1 ml Glacial acetic acid Gentian Violet 1 ml 100 ml distilled water

Filter before use

Dried Blood

Spread blood on glass plate. Dry in oven or in air.

When dry scrape into beaker.

Drosophila Chromosome Stain #1

Synthetic Orcein 2 gm 50 ml Glacial acetic acid 50 ml 85% Lactic acid

Dissolve Orcein in hot glacial acetic acid, remove from heat and add the Lactic acid. Filter.

Drosophila Chromosome Stain #2

1 gm Synthetic Orcein 45 ml Glacial acetic acid

25 ml 85% Lactic acid 30 ml distilled water

Dissolve Orcein in hot glacial acetic acid, remove from

heat and add the water and Lactic acid. Filter.

F. ' S

<u>A B C D E F G H I J K L M N O P Q R S T U V W X Y Z</u>

Eosin (microtechnique)

0.5gm Eosin 100 ml 95% Ethanol

Erlich Solution for Indol Test

1 gm Para dimethylamino

benzaldehyde 95% ethanol

95 ml 95% ethano 20 ml con HCl

Dissolve the powder in the alcohol. Then add the HCl and

mix well

F's

<u>ABCDEFGHI_JKLMNOPQRSTUVWXYZ</u>

F.A.A. (Carl's Solution)

170 ml 95% ethanol 60 ml Formalin

20 ml Glacial acetic acid

280 ml water

Add the glacial acetic acid to the solution just

before using.

F.A.A. Alexander's

120 ml Formalin 300 ml 95% Ethanol

20 ml Glacial acetic acid

600 ml tap water

F.A.A., Lavdowsky's

20 ml Formalin 100 ml Ethanol

4 ml Glacial acetic acid 80 ml distilled water

F.A.A., Botanical

115 ml Formalin

115 ml Glacial acetic acid 805 ml 95% Ethanol 805 ml Distilled water *****************

F.A.A., Smith's

90 ml 85% isopropanol 10 ml glacial acetic acid

10 ml Formalin

Farmers fluid

100% ethanol 3 parts

1 part Glacial acetic acid

Fixative, Navashin's

SOLUTION A

5 gm Chromic acid 50 ml Acetic acid 320 ml distilled water

SOLUTION B

200 ml formalin 175 ml distilled water

Mix equal parts of each solution just before use.

Fixative, Transeau's

supernatant of sat 6 parts

aq. CuSo4 95% ethanol 3 parts 40% aq. formalin 1 part

Used to preserve color in plants

Formalin, Neutral buffered, Lillie's

100 ml formalin 900 ml distilled water 4 g NaH2-P04

Na2-HPO4 6.5 gm

Frog Ringers

0.8 gm NaCl

0.02 gmCaCl2 anhydrous

0.02 gm KCl 0.02 gm NaHCO3 100 ml distilled water

** 0.1 gm Dextrose

** add only if requested and if requested add just before

use.

FRUIT FLY MEDIA

BREWERS YEAST 37 GM 150 GM CORN MEAL 35 GM AGAR

200 ML

UNSULFURED MOLASSAS 3 GM MOLD INHIBITOR (CAROLINA BIOLOGICAL

1900 ML DISTILLED WATER

Mix yeast, mold inhibitor, and agar with a little 95% ethanol. Add some of the water to the corn meal to wet it and then add all the ingredients to the remaining water. While stirring constantly bring the mixture to a boil. Dispense in 50 ml alaquats in 1 pint milk bottles.

G's

<u>ABCDEFGH_IJKLMNOPQRSTUVWXYZ</u>

Gastric Juice or enzyme

1-2 gm pepsin 100 ml 0.2% HCl

0.2% HCl = 6 ml con HCl in 1 liter of distilled water

Geimsa Stain

Dissolve 0.5 gm of powder in 33 ml of glycerine at 55 to 66 deg. C overnight with stirring and heat. To this add 33 ml of methyl alcohol.
WORKING DILLUTION =

1 ml of above stain to 25 ml of distilled water.

Gentian Violen Stain

1 gm Gentian Violet 20 gm Ethanol 80 ml 5% Formalin

Dissolve the Gentian Violet in the alcohol then add the

formalin, mix and filter.

Gilson's Mixture

15 ml Nitric acid sp =1.45

or 70-80 %

4 ml Glacial acetic acid
20 gm Mercuric chloride
100 ml 60% ethanol
880 ml distilled water

Can be heated to dissolve but let sit for a day or so to let it pcppt out. Do not use metal probes as the mercuric chloride eats metal and will cause specimen to stick to

probe.

Store in clear glass container.

Glycerine Alcohol

90 parts 70% ethanol 10 parts Glycerine

Glycerine Jelly

10 gm gelatin

60 ml distilled water
70 ml Glycerine
1 gm Phenol

Dissolve phenol in water then stir in the glycerine. When miced, add gelatin and heat until dissolved. Transfer to a

widemouth jar and allow to cool.

Glycerine Preservative for Kelp

3 parts Water
1 part Glycerine
10 gm/L Phenol

Add kelp, cleaned of sand and epiphytes, making sure they are completly immersed (a weight may be added if necessary) and preserve for about 10 days. Remove and dry on newapaper 10 - 14 days. Turn occasionally so that all portions will be dry. Glycerine solution may be used several times.

Gower's R-B-C

12.2 gm Sodium Sulfate
33.3 ml Glacial acetic acid
200 ml distilled water

Grenacher's Alcoholic Borax Carmine

3 gm Carmine 4 gm Borax

100 ml distilled water

Boil until the carmine is dissolved (1/2 hr) or better yet allow the mixture to stand at room temp until dissolved. Then add 70% ethanol (100 ml) and allow to stand for a day

or two then filter

Grenacher's Alum Carmine

100 ml distilled water 3 gm aluminum ammonium

sulfate 1 gm Carmine

Combine and boil for 15 min or until carmine is dissolved.

Cool and filter.

H's

$\underline{A} \quad \underline{B} \quad \underline{C} \quad \underline{D} \quad \underline{E} \quad \underline{F} \quad \underline{G} \quad \underline{H} \quad \underline{I} \quad \underline{J} \quad \underline{K} \quad \underline{L} \quad \underline{M} \quad \underline{N} \quad \underline{O} \quad \underline{P} \quad Q \quad \underline{R} \quad \underline{S} \quad \underline{T} \quad U \quad V \quad \underline{W} \quad X \quad \underline{Y} \quad \underline{Z}$

Hagerup's Fixative

Harris' Alum Hematoxylin

0.5 gm Hematoxylin 20 gm Aluminum Ammonium

Sulfate

100 ml distilled water 0.5gm Mercuric Oxide

Boil the hematoxylin and ammonium alum together in distilled water (use a large container). Add the mercuric oxide with care as there is a tendency for the solution to boil

up.

Helly's Fluid

2.5 gm Potassium Dichromate 5.0 gm Mercuric Chloride 1.0 gm Sodium sulfate 100 ml distilled water

5 ml Formalin

Hematoxylin, Harris'

0.5 gm Hematoxylin 20 gm Aluminum Ammonium

Sulfate

100 ml distilled water 0.5gm Mercuric Oxide

Boil the hematoxylin and ammonium alum together in distilled water (use a large container). Add the mercuric oxide

with care as there is a tendency for the solution to boil ***************** Hematoxylin, Iron 2 solutions are used. They are not to be mixed. 1. Ferric alum (clear violet crystals) 2.5 gm (ferric ammonium sulfate) is dissolved in 100 ml of distilled water. 2. 5 gm of Hematoxylin is dissolved in 1 liter of distilled water and 100 ml of 95% ethanol. ***************** Holtfreter's Solution 0.35 gm NaCl $0.005 \, \mathrm{gm}$ KCl CaCl2 0.01qm0.02gmNaHCO3 100 ml distilled water ***************** Honey Peptone Agar 10 gm Peptone 60 gm Honey 15 gm agar distilled water 1 Liter ***************** Houpt's Adhesive 1 gm of plain Knox gelatin (or reagent gelatin) is dissolved in 100 ml of water at 30 deg C. Add 2 gm phenol and 15 ml of glycerine. Stir and filter (takes 24 hrs). Hoyer's Medium 25 ml water 15 gm Gum Arabic 10 ml Glycerine 100 gm Chloral hydrate First add crystal of chloral hydrate the size of a pea to prevent fungal growth. Then soak gum-water-pea of chloral hydrate for 24 hrs. Then add 100 gm of chloral Hydrate and wait until dissolved. This may take several days. After everything is dissolved add glycerine. ***************** Hydrochloric acid dillutions 1 N 86 mls to volume of 1 liter 2 N 172 mls to volume of 1 liter 3 N 258 ml to volume of 1 liter ***************** HYDROCHLORIC ACID DILLUTIONS 258ml/L 36% 3N **************** Hydroxide Sodium Iodide for Aquatic Analysis Add 100 grams of NaOH and 27 grams of NaI to 1 liter of water. ***************** I's <u>ABCDEFGHIJKLMNOPQRSTUVWXYZ</u> Saturday, January 13, 2001 Recipies Page: 19

Indicator, Methyl Orange

0.5 gm Methyl orange

1 liter water

Indicators, pH

Brom Cresol green; $0.4~\mathrm{gm}$ BCG, 500 ml 95% ethanol, 500 ml distilled water. Filter.

Cresol Red; Same procedure as above using 0.2gm cresol red Thymol blue; Same as above using 0.5 gm of thymol blue Phenolphthalein; 1.0 gm phenolphthalein in 50 ml of 95% ethanol and 50 ml of distilled water.

Indol Acetic Acid in Lanolin

5 gm lanolin 2 mg IAA

Warm (do not boil) the lanolin in a small screw cap vial until the lanolin just turns to liquid. Add the IAA and mix thoroughly with a glass rod and allow to cool.

Indol acetic acid solution

Indol acetic acid must first be dissolved in a small amount of 95% ethanol before adding the appropriate amount of water to reach the desired concentration

NOTE: IAA is unstable as a solution and bust be refrigerate Even cold it will only be good for about 36 hours.

Indol Test (Kobacs' Reagent)

75 ml Amyl alcohol 25 ml con HCl

5 gm paradimethylamino-

benzaldehyde

Should be made up in small quantities and stored in refrigerator in clear glass stoppered bottles.

Insect Saline

0.7% NaCl 0.2 ml 10% CaCl2

Insect Saline (Hayome)

0.7% NaCl in distilled

water.

Iodine - Reagent to devolatalize Iodine

0.8 gm NaOH

0.5 gm Sodium thiosulfate
0.3 gm Potassoum Iodide
to 1 liter distilled water

Iodine solution (LUGOL'S)

50 gm Iodine

100 gm KI

to 1 liter distilled water

Mix I and KI in a mortar. Add distilled water in small portions to wash out the contents into a graduated cylinder.

Iodine Solution (STOCK for starch)

35 gm KI 5 gm I

500 ml distilled water

for working dillution to use in starch test dilltue 1 part

stock with 2 parts water.

K's

A B C D E F G H I J K L M N O P Q R S T U V W X Y Z

K.A.S.A.

158 ml crude Kerosene
473.75 ml 95% isopropanol
105.25 ml glacial acetic acid
263 ml sec-butyl alcohol
Add sec-butyl alcohol last until solution clears

Karpaenchenko's fixative (Taylor's modification)

1.5ml 10% ag chromic acid

10 ml 10% aq acetic acid

0.83 ml formalin 23.67 ml distilled water

Knop's solution pH 7.6

1 gm MgSO4*7HOH 0.20 gm KH2-PO4 1.0 gm KNO3 1 gm Ca(NO3)2*4HOH

Add each of the above to 250~ml of distilled water then combine to 1 liter adding Ca Nitrate last. Add 1 drop

of 1% ferric chloride (freshly prepared)

Knops Solution (acid medium pH aprox 6.4)

0.25 gm MgSO4*7HOH 0.25 gm KH2-PO4 0.12 gm KCl

1.0 gm Ca(NO3)2*4HOH

Add each of the above to 250~ml of distilled water then combine to 1 liter adding Ca Nitrate last. Add 1 drop

of 1% ferric chloride (freshly prepared)

Kobacs' Reagent

75 ml Amyl alcohol 25 ml con HCl

5 gm paradimethylaminobenzaldehyde

Should be made up in small quantities and stored in refrigerator in clear glass stoppered bottles.

Kovacs' Reagent - for indol test

75 ml Amyl alcohol 25 ml con HCl

5 gm Paradimethylaminobenzaldehyde

Should be made up in small quantities and stored in refrig

in clear glass, stoppered bottles.

Kreb's Manometer Fluid

44 gm NaBr anhydrous
0.3 gm Triton X-100
0.3 gm Evans blue
1 liter distilled water

Density = 1.033 at 20 deg C.

Kymograph Shellac Solution

Dillute 1 volume of shellac with 2.5 volumes of 95% ethanol.

L's

<u>A B C D E F G H I J K L M N O P Q R S T U V W X Y Z</u>

Lactophenol Mounting medium

Langeron's recipie calls for 2 parts glycerine to 1 part

of the other reagents.

Lavdowsky's fixative

20 ml Formalin 100 ml Ethanol

4 ml Glacial acetic acid 80 ml distilled water

Litmus solution

Add powdered litmus to distilled water to make a dark

blue solution

Locke's Physiological Saline

9 gm NaCl
0.2 gm NaHCO3
0.42 gm KCl
0.25 gm CaCl2
2 gm dextrose
1000ml distilled water

Dissolve CaCl2 in a little of the water and add it to the

rest of the solution to prevent poptation.

Add dextrose only if specified, and then at the last moment.

Loeffler's methylene blue

0.3 gm Methylene blue 30 ml 95% ethanol

70 ml Dilute KOH (0.017%)

LUGOL'S iodine solution

50 gm Iodine 100 gm КT

distilled water to 1 liter

Mix I and KI in a mortar. Add distilled water in small portions to wash out the contents into a graduated cylinder.

M's

<u>ABCDEFGHIJKLMNOPQRSTUVWXYZ</u>

Maceration Fluid

1 part 10% Nitric acid 1 part 10% Chromic acid (chromic trioxide)

Boil material 1 min in water. Pour off water add acid, stir with glass rod and check at intervals until it begins to seperate. Pour off acid, put in white silk scarf or handkerchief and rinse with water for 12 hours. Drain off water and cover with 5 volumes of 70% ethanol.

Mallory's Triple stain

First Stain Solution; 1% acid fuchsin Differentiating Moderating solution; 1% Phosphotungstinic acid.

Second Staining Solution;

100 ml water

0.5 gm Analine blue

2.0 gm Orange G

2 gm Oxalic Acid

Manometer Fluid, Brodie's

23 gm

Sodium Choleate 5 gm 500 ml distilled water

density = 1.033, Po = 10000

Evans blue (200mg/L) or acid fuchsin are excellent dyes for the fluid; other dyes may be used, but some of these tend to decompose in the manometer. Determine the density

of the solution with a pycnometer.

Manometer Fluid, Kreb's

44 gm NaBr anhydrous 0.3 gm Triton X-100 0.3 gm Evans blue 1 liter distilled water

Density = 1.033 at 20 deg C.

Mayer's Paracarmine

1 gm Carminic acid 0.5 gm Al chloride CaCl2 4.0 gm 70% ethanol 100 ml

1.5 gm KI 5 gm I 20 ml water

20 gm Chloral Hydrate

Methocel

2.5-3 gm Methocel 90 ml distilled water

Mic methocel with the 90 ml of Hot not boiling water. Cool to 5 deg C (in refrigerator). It should be transparent when

finished.

Methyl Green

1 gm Methyl green 1 ml glacial acetic acid 99 ml water

Dissolve satin in acid and dilute with water

Methyl Green Glycerine-Jelly Stain

Add saturated (about 3%) solution of methyl green in 50% ethanol drop by drop to melted glycerine jelly (a good comercial grade is satisfactory), until jelly becomes as dark as green ink.

Methyl Orange Indicator

0.5 gm Methyl orange

1 liter water

Methylene blue stain

0.3 gm Methylene blue 30 ml 95% ethanol 100 ml distilled water

Dissolve stain in alcohol, then dilute with water and filter

Methylene Blue, Loeffler's

0.3 gm Methylene blue 30 ml 95% ethanol 70 ml Dilute KOH

Meyer's Adhesive

Add to one fresh egg white, an equal quantity of glycerine and 1 gm of sodium salicylate and a crushed crystal of thymol. Shake wall and filter through 2-3 thicknesses of sterile cheesecloth

N's

 $\underline{A} \quad \underline{B} \quad \underline{C} \quad \underline{D} \quad \underline{E} \quad \underline{F} \quad \underline{G} \quad \underline{H} \quad \underline{I} \quad \underline{J} \quad \underline{K} \quad \underline{L} \quad \underline{M} \quad \underline{N} \quad \underline{O} \quad \underline{P} \quad \underline{Q} \quad \underline{R} \quad \underline{S} \quad \underline{T} \quad \underline{U} \quad \underline{V} \quad \underline{W} \quad \underline{X} \quad \underline{Y} \quad \underline{Z}$

NADI Reagent

A. 1% alpha-naphthol in 95% ethanol.

B. 1% N,N-dimethyl-p-phenylenediamine HCl in water Solutions a and b are mixed in equal amounts just prior to

Naphthylamine Hydrochloride

Dissolve $0.6~\rm gm$ 1-naphthylamine hydrochloride in distilled water to which $1.0~\rm ml$ of con HCl has been added. Dilute to $100~\rm ml$ with distilled water. Store in refrigerator.

Navashin's Fixative

SOLUTION A

5 gm Chromic acid 50 ml Acetic acid 320 ml distilled water

SOLUTION B

200 ml formalin 175 ml distilled water

Mix equal parts of each solution just before use.

Nessler Reagent

Dissolve 100 gm anhy7drous mercuric iodide and 70 gm anhydrous potassoum iodide in a small quantity of water and add mixture slowly to a cool solution of 160 gm of NaOH in 500 ml of distilled water. Dilute to 1 liter with distilled water. Store in pyrex bottle out of direct sunlight.

Neutral Buffered Formalin, Lillie's

100 ml formalin 900 ml distilled water

4 g NaH2-P04 6.5 gm Na2-HP04

Newcomers Fixative

6 parts isopropyl alcohol
3 parts propionic acid
1 part petrolium ether

1 part acetone
1 part dioxane

Ninhydrin Reagent

0.5% ninhydrin in acetone or n-bunanol. Thomas prefers n-butanol.

Nitric acid dillution

3N 195 ml/L 69%

0's

 $\underline{A} \ \underline{B} \ \underline{C} \ \underline{D} \ \underline{E} \ \underline{F} \ \underline{G} \ \underline{H} \ \underline{I} \ \underline{J} \ \underline{K} \ \underline{L} \ \underline{M} \ \underline{N} \ \underline{O} \ \underline{P} \ \underline{Q} \ \underline{R} \ \underline{S} \ \underline{T} \ \underline{U} \ \underline{V} \ \underline{W} \ \underline{X} \ \underline{Y} \ \underline{Z}$

Ortho-Toluidine Reagent (for aquatic analysis)

Dissolve 4 gm ortho-toluidine in distilled water containing 15 ml of con HCl. Dilute to a final volume of 500 ml.

Pancreatic juice, artificial

40 gm Pancreatin
0.4 gm Baking soda

200 ml water

Mix in diatomaceous earth and then filter under suction Add a few drops of chloroform. Incubate at body temperature Unstable. Maybe kept overnight in refrigerator.

P's

A B C D E F G H I J K L M N O P Q R S T U V W X Y Z

Paradon (cellodine)

1:1 !00 % paradon and di-ethyl ether

Pepsin in HCl

10-2 gm Pepsin 100 ml 0.2% HCl

Pepsin Solution (make fresh)

1-2 gm Pepsin 100 ml water

Peroxides Test

Add 1 ml of 10% KI to 10 ml of solution to be tested. If brown color results in either layer peroxides are present and the solution should be discarded with proper precautions.

pH indicators

Brom Cresol green; $0.4~\mathrm{gm}$ BCG, 500 ml 95% ethanol, 500 ml distilled water. Filter.

Cresol Red; Same procedure as above using 0.2gm cresol red Thymol blue; Same as above using 0.5 gm of thymol blue Phenolphthalein; 1.0 gm phenolphthalein in 50 ml of 95% ethanol and 50 ml of distilled water.

Phenol Cotton Blue

20 ml Carbolic acid
20 ml Lactic acid
40 ml Glycerine
20 ml Water
0.05 gm Cotton blue

Phenol Red Lactose Broth

10 gm Protease Peptone #3

1 gm Beef extract 5 gm Lactose

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5 gm Sodium Chloride 18 mg Phenol red 1 liter water

Phenoldisulfonic acid

Dissolve 25 gm pure white phenol crystals in 150 ml con Sulfuric acid. Add 75 ml fuming sulfuric acid (15% free SO4 Stir well and heat for 2 hours in water bath

Phenolphthalein

0.5 gm Phenolphthalein 100 ml 50% ethanol

Phloroglucinol, HCl

Solution A; 1 gm phloroglucinol in 100 ml of 95% ethanol Solution B; 50 ml con HCl in 50 ml distilled water

Phosphoric acid dilltuion

3 N 205 m/L 85%

Physiological saline, LOCKE'S

9 gm NaCl
0.2 gm NaHCO3
0.42 gm KCl
0.25 gm CaCl2
2 gm dextrose
1000ml distilled water

Dissolve CaCl2 in a little of the water and add it to the

rest of the solution to prevent poptation.

Add dextrose only if specified, and then at the last moment.

Plant maceration fluid

Boil material 1 min in water. Pour off water add acid, stir with glass rod and check at intervals until it begins to seperate. Pour off acid, put in white silk scarf or handkerchief and rinse with water for 12 hours. Drain off water and cover with 5 volumes of 70% ethanol.

Pollen Acetylizing Mixture

9 parts Acetic Anhydride 1 part con Sulfuric Acid

Pollen Mounting Medium, Dahl's

16 parts 85% Lactic acid 2 parts N-Butyl alcohol 1 part 95% Ethanol

Saturday, January 13, 2001 Recipies 1 part Glacial Acetic Acid 1 part Glycerine ************* Pollen Trapping Oil 2 parts melted white petrolatum 1 part Parafin oil 1 part butyl alcohol 1 part xylene Mix in beaker on hot blate. Filter while warm; keep in stoppered containers, free from contamination. ***************** Potassium Oxalate - Aquatic analysis tests for O2 20 gm K2-C2-O4*HOH dissolved in water Add 45 gm NaOH and diltue to 1 liter with water. ***************** Prescott's Preservative for Green Plant Material 6 parts water 95% ethanol 3 parts 1 part Formalin 5 ml/100ml Glycerine Saturated with CuSO4 - decant leaving excess CuSO4 ****************** Preservative (Sutton's) 10% glycerine 3% Phenol 2% Formalin 85% Water ***************** Pseudomonas Ovalic Culture Medium 1% Yeast extract 1% Beef extract 1.8% agar ***************** Pyrogallic Acid 50 mg KOH 100 ml distilled water

Pyrogallol

<u>ABCDEFGHIJKLMNOPQRSTUVWXYZ</u>

Rat Ringers

0.9% NaCl 0.24% CaCl2 0.042% KC1 0.02% NaHCO3 0.1% Dextrose (if requested) *********

Rat Ringers, LOCKE'S

0.9% NaCl

0.24% CaCl2 0.042% KCl 0.02% NaHCO3 0.1% Dextrose

Rennin Solution

1-2 gm Rennin 100 ml water

Riboflavin Stock

Dissolve 5 mg of Riboflavin in 30 ml of 0.02N acetic acid. Warm in a water bath with constant rotation until dissolved. Cool to room temp and dilute to 100 ml by adding additional 0.02N acetic acid. Store refrigerated.

Standard - unstable

10 ml of stock brought to 500 ml with distilled water. Working; Dilute 1:10 before using.

Ringers for lower vertebrates

0.65% 0.02% KCl CaCl2 0.02% 0.02% NaHCO3

Ringers, Frog

0.8 gm NaCl

0.02 gmCaCl2 anhydrous

0.02 gm KCl 0.02 gm NaHCO3

100 ml distilled water

** 0.1 gm Dextrose

** add only if requested and if requested add just before

Ringers, Rat

0.9% NaCl 0.24% CaCl2 0.042% KC1 0.02% NaHCO3 0.1% Dextrose (if requested)

S's

<u>ABCDEFGHIJKLMNOPQRSTUVWXYZ</u>

Safranin for plant material

Dissolve 1 qm of Safranin O in 50 ml of methyl cellosolve.

After dissolved add:

95% ethanol 25 ml distilled water 25 ml Sodium acetate 1 gm Formalin 2 ml

Saline, Insect

0.7% NaCl 0.2 ml 10% CaCl2

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Saline, Insect (Hayome)

0.7% NaCl in distilled

Saline, Physiological, Locke's

9 gm NaCl
0.2 gm NaHCO3
0.42 gm KCl
0.25 gm CaCl2
2 gm dextrose
1000ml distilled water

Dissolve CaCl2 in a little of the water and add it to the

rest of the solution to prevent poptation.

Add dextrose only if specified, and then at the last moment.

SCHIFF'S REAGENT

5 gm Basic Fuchsin

1N HCl 100 ml K2-S2-O5 10 gm

activated charcoal

(norite) 5 gm

Disolve 5 grams of basic fuchsin in 900 ml boiling distilled water. Cool to 50 degrees C and add 100 ml of 1N HCl cool to 25 deg and then add 10 grams of K2S2O5. Shake 2 minutes and leave in the dark for 24 hours. Add 5 grams of fine activated charcoal, shake for 2 minutes and then filter through fast paper. Filtrate should be clear. Store in a foil covered bottle in the refrigerator

Schoudinn's Fluid

33 ml

66 ml sat aq. Mercuric

chloride 95% ethanol

5 ml glacial acetic acid

Add acid just prior to use.

Semicon's Carmine

1 part glacial acetic acid

1 part water

Add carmine powder in excess of that which dissolves immediately. Heat to 95-100 deg C for 15 min. Cool and filter. Dilute filtrate eith equal partsw of 70% ethanol

Shellac solution for Kymographs

Dillute 1 volume of shellac with 2.5 volumes of 95% ethanol.

Smith's F.A.A.

90 ml 85% isopropanol 10 ml glacial acetic acid

10 ml Formalin

Soil Extract Agar

960 mls Bristols solution 40 ml soil water supnatant

15 gm agar

Soil water Medium

In one gallon battery jar place 0.5 gm calcium carbonate enough garden soil to make a layer aprox 1/2 to 3/4 inch deep and distilled water to within one inch of the top. Cover with a glass plate and steam for 1 hour on 2 consecutive days.

Spindle Inhibitor in Mitosis

0.01-0.05% Colchicine 4-6hrs sat aq Paradichlorobenzene

4 hrs

sat aq alpha bromonapthalen

4 hrs

Starch solutions

Make a thin slurry with the starch and a small amount of the water. Heat the rest of the water to boiling. Add starch slurry while stirring bring back to a boil. Cool before use. 0.1% final concentration works best for iodine test

Sudan Black B

0.25 gm Sudan Black B 100 ml Ethylene glycol

Dissolves slowly - let stand overnight and filter.

Very viscous.

Sudan IV

Saturate sudan IV in absolute ethanol.

Sulfanilic Acid

Completely dissolve 0.60 gm sulfanilic acid in 70 ml of hot distilled water. Cool the solution and add 20 ml con HCl. Dilute to 100 ml with distilled water and mix thoroughly.

Sulfuric acid dillutions

3N 168 ml/L 95%

Sutton's Preservative

10% Glycerine
3% Phenol
2% Formalin
85% Water

Sutton's preservative

10% glycerine
3% Phenol
2% Formalin
85% Water

T's

3 parts

1 part

$\underline{A} \quad \underline{B} \quad \underline{C} \quad \underline{D} \quad \underline{E} \quad \underline{F} \quad \underline{G} \quad \underline{H} \quad \underline{I} \quad J \quad \underline{K} \quad \underline{L} \quad \underline{M} \quad \underline{N} \quad O \quad \underline{P} \quad Q \quad \underline{R} \quad \underline{S} \quad \underline{T} \quad U \quad V \quad \underline{W} \quad X \quad \underline{Y} \quad \underline{Z}$

Transeau's Fixative

6 parts supernatant of sat

aq. CuSo4 95% ethanol 40% aq. formalin

Trichrome Stain

0.6 gm Chromotrope 2R
0.3 gm Fast green FCF
0.7 gm Phosphotungstic acid
1 ml Glacial acetic acid
100 ml distilled water

Triple Stain, Mallory's

First Stain Solution; 1% acid fuchsin

Differentiating Moderating solution; 1% Phosphotungstinic

acid.

Second Staining Solution;

100 ml water

0.5 gm Analine blue

2.0 gm Orange G

2 gm Oxalic Acid

Tris-Maleate buffer

Dissolve 29 gm of maleic acid and 30.3 gm of tris-hydroxymethyl aminomethane in 500 ml of water. Add 2 g of Charcoal, shake and let stand for 10 min. Filter. Mix 40 ml of this stock solution with the amount of 1N NaOH indicated below and dillute to a final volume of 100 ml using water.

Turk's Diluting Fluid

1 ml Glacial acetic acid 1 ml Gentian Violet 100 ml distilled water

Filter before use

W's

 $\underline{\mathbf{A}} \quad \underline{\mathbf{B}} \quad \underline{\mathbf{C}} \quad \underline{\mathbf{D}} \quad \underline{\mathbf{E}} \quad \underline{\mathbf{F}} \quad \underline{\mathbf{G}} \quad \underline{\mathbf{H}} \quad \underline{\mathbf{I}} \quad \underline{\mathbf{J}} \quad \underline{\mathbf{K}} \quad \underline{\mathbf{L}} \quad \underline{\mathbf{M}} \quad \underline{\mathbf{N}} \quad \underline{\mathbf{O}} \quad \underline{\mathbf{P}} \quad \underline{\mathbf{Q}} \quad \underline{\mathbf{R}} \quad \underline{\mathbf{S}} \quad \underline{\mathbf{T}} \quad \mathbf{U} \quad \underline{\mathbf{V}} \quad \underline{\mathbf{W}} \quad \mathbf{X} \quad \underline{\mathbf{Y}} \quad \underline{\mathbf{Z}}$

Waris Solution

To 1 liter of distilled water add 1 ml of the following: 10% KNO3, 2% MgSO4*7HOH, 2% (NH4)2-HPO4

5% CaSO4,

Iron sequestrin solution (adjust to pH 6.0 with 0.01N HCl

or 0.01N KOH

= 2.61 gm Sequestrin AA 2.49 gm FeSO4*7HOH

27 ml 1 N KOH

500 ml distilled water

White blood cell dilution fluid

1% Glacial acetic acid

White's Basic Medium

0.1 gm Ca(NO3)2

0.08 gm KNO3 0.035 gm MgS04 0.065 gm KCl KH2-PO4 0.0125 0.00075 gm ΚI MnSO4 0.0044 gm0.0025 gmFe2-(SO4)3 0.0015 gm ZnSO4 0.0016 gmH3-B04 20 gm Sucrose Bring to 1 liter with distilled water.

Y's

$\underline{A} \quad \underline{B} \quad \underline{C} \quad \underline{D} \quad \underline{E} \quad \underline{F} \quad \underline{G} \quad \underline{H} \quad \underline{I} \quad J \quad \underline{K} \quad \underline{L} \quad \underline{M} \quad \underline{N} \quad O \quad \underline{P} \quad Q \quad \underline{R} \quad \underline{S} \quad \underline{T} \quad U \quad V \quad \underline{W} \quad X \quad \underline{Y} \quad \underline{Z}$

Yeast in Congo Red

1 pkg dry yeast 0.25-0.5gm Congo red water

Add yeast to warm water and stir in congo red. Let stand all day. Heat to boiling and cool. Store in refrigerator.

Z's

<u>A B C D E F G H I J K L M N O P Q R S T U V W X Y Z</u>

Ziehl-Nielsen's Carbolfuchsin - Acid Fast

0.3 gm Basic fuchsin

90% dye content 95% ethanol

10 ml 95% eth 5 gm Phenol

95 ml distilled water

Dissolve fuchsin in alcohol. Dissolve Phenol in water and

then mix solutions.